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1. Your reference — SMW/CP5236674

2. Patent application number *(The Patent Office will fill in this part)* — 24 MAY 1996 **9610967.3**

3. Full name, address and postcode of the or of each applicant *(underline all surnames)*

CAMBRIDGE ANTIBODY TECHNOLOGY LIMITED
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Patents ADP number *(if you know it)* 5781972002

If the applicant is a corporate body, give the country/state of its incorporation — UNITED KINGDOM

4. Title of the invention — SPECIFIC BINDING MEMBERS, MATERIALS AND METHODS

5. Name of your agent *(if you have one)* ~~MEWBURN ELLIS~~

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Country	Priority application number *(if you know it)*	Date of filing *(day / month / year)*

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Number of earlier application	Date of filing *(day / month /year)*

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12. Name and daytime telephone number of person to contact in the United Kingdom — Seán M. Walton — 0171 240 4405

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SPECIFIC BINDING MEMBERS, MATERIALS AND METHODS

This invention relates to specific binding members for a foetal isoform of fibronectin, ED-B, which is also expressed in the developing neovasculature of tumours, as demonstrated both by immunocytochemistry and by targeting of tumours *in vivo*. It also relates to materials and methods relating to such specific binding members.

The primary aim of most existing forms of tumour therapy is to kill as many constituent cells of the tumour as possible. The limited success that has been experienced with chemotherapy and radiotherapy relates to the relative lack of specificity of the treatment and the tendency to toxic side-effects on normal tissues. One way that the tumour selectivity of therapy may be improved is to deliver the agent to the tumour through a binding protein, usually comprising a binding domain of an antibody, with specificity for a marker antigen expressed on the surface of the tumour but absent from normal cells. This form of targeted therapy, loosely termed 'magic bullets', has been mainly exemplified by monoclonal antibodies (mAbs) from rodents which are specific for so-called tumour-associated antigens expressed on the cell surface. Such mAbs may be either chemically conjugated to the cytotoxic moiety (for example, a toxin or a drug) or may be produced as a

recombinant fusion protein, where the genes encoding the mAb and the toxin are linked together and expressed in tandem.

The 'magic bullet' approach has had limited, although significant, effect in the treatment of human cancer, most markedly in targeting tumours of lymphoid origin, where the malignant cells are most freely accessible to the therapeutic dose in the circulation. However, the treatment of solid tumours remains a serious clinical problem, in that only a minute proportion of the total cell mass, predominantly the cells at the outermost periphery of the tumour, is exposed to therapeutic immunoconjugates in the circulation; these peripheral targets form a so-called 'binding site barrier' to the tumour interior (Juweid et al, 1992, *Cancer Res.* 52 5144-5153). Within the tumour, the tissue architecture is generally too dense with fibrous stroma and closely packed tumour cells to allow the penetration of molecules in the size range of antibodies. Moreover, tumours are known to have an elevated interstitial pressure owing to the lack of lymphatic drainage, which also impedes the influx of exogenous molecules. For a recent review of the factors affecting the uptake of therapeutic agents into tumours, see Jain, R (1994), *Sci. Am.* 271 58-65.

Although there are obvious limitations to treating

solid tumours through the targeting of tumour-associated antigens, these tumours do have a feature in common which provides an alternative antigenic target for antibody therapy. Once they have grown beyond a certain size, tumours are universally dependent upon an independent blood supply for adequate oxygen and nutrients to sustain growth. If this blood supply can be interfered with or occluded, there is realistic potential to starve thousands of tumour cells in the process. As a tumour develops, it undergoes a switch to an angiogenic phenotype, producing a diverse array of angiogenic factors which act upon neighbouring capillary endothelial cells, inducing them to proliferate and migrate. The structure of these newly-formed blood vessels is highly disorganised, with blind endings and fenestrations leading to increased leakiness, in marked contrast to the ordered structure of capillaries in normal tissue. Induction of angiogenesis is accompanied by the upregulation of expression of certain cell surface antigens, many of which are common to the vasculature of normal tissues. Identifying antigens which are unique to neovasculature of tumours has been the main limiting factor in developing a generic treatment for solid tumours through vascular targeting. The antigen which is the subject of the present invention addresses this problem directly.

During tumour progression, the extracellular matrix of the surrounding tissue is remodelled through two main processes: (1) the proteolytic degradation of extracellular matrix components of normal tissue and (2) the *de novo* synthesis of extracellular matrix components by both tumour cells and by stromal cells activated by tumour-induced cytokines. These two processes, at steady state, generate a 'tumoral extracellular matrix', which provides a more suitable environment for tumour progression and is qualitatively and quantitatively distinct from that of normal tissues. Among the components of this matrix are the large isoforms of tenascin and fibronectin (FN); the description of these proteins as isoforms recognises their extensive structural heterogeneity which is brought about at the transcriptional, post-transcriptional and post-translational level (see below). It is one of the isoforms of fibronectin, the so-called B+ isoform, that is the subject of the present invention.

Fibronectins (FN) are multifunctional, high molecular weight glycoprotein constituents of both extracellular matrix and body fluids. They are involved in many different biological processes such as the establishment and maintenance of normal cell morphology, cell migration, haemostasis and thrombosis, wound healing and oncogenic transformation (for reviews see

Alitalo et al., 1982; Yamada, 1983; Hynes, 1985; Ruoslahti et al., 1988; Hynes, 1990; Owens et al., 1986). Structural diversity in FNs is brought about by alternative splicing of three regions (ED-A, ED-B and IIICS) of the primary FN transcript (Hynes, 1985; Zardi et al., 1987) to generate at least 20 different isoforms, some of which are differentially expressed in tumour and normal tissue. As well as being regulated in a tissue- and developmentally-specific manner, it is known that the splicing pattern of FN-pre-mRNA is deregulated in transformed cells and in malignancies (Castellani et al., 1986; Borsi et al, 1987; Vartio et al., 1987, Zardi et al, 1987; Barone et al, 1989; Carnemolla et al, 1989; Oyama et al, 1989, 1990; Borsi et al, 1992b). In fact, the FN isoforms containing the ED-A, ED-B and IIICS sequences are expressed to a greater extent in transformed and malignant tumour cells than in normal cells. In particular, the FN isoform containing the ED-B sequence (B+ isoform), is highly expressed in foetal and tumour tissues as well as during wound healing, but restricted in expression in normal adult tissues (Norton et al, 1987; Schwarzbauer et al, 1987; Gutman and Kornblihtt, 1987; Carnemolla et al, 1989; ffrench-Constant et al, 1989; ffrench-Constant and Hynes, 1989; Laitinen et al, 1991.) B+ FN molecules are undetectable in mature vessels, but upregulated in

angiogenic blood vessels in normal (e.g. development of the endometrium), pathologic (e.g. in diabetic retinopathy) and tumour development (Castellani et al, 1994).

The ED-B sequence is a complete type III-homology repeat encoded by a single exon and comprising 91 amino acids. In contrast to the alternatively spliced IIICS isoform, which contains a cell type-specific binding site, the biological function of the A+ and B+ isoforms is still a matter of speculation (Humphries et al., 1986).

The presence of B+ isoform itself constitutes a tumour-induced neoantigen, but in addition, ED-B expression exposes a normally cryptic antigen within the type III repeat 7 (preceding ED-B); since this epitope is not exposed in FN molecules lacking ED-B, it follows that ED-B expression induces the expression of neoantigens both directly and indirectly. This cryptic antigenic site forms the target of a monoclonal antibody (mAb) named BC-1 (Carnemolla et al, 1992). The specificity and biological properties of this mAb have been described in EP 0 344 134 B1 and it is obtainable from the hybridoma deposited at the European Collection of Animal Cell Cultures, Porton Down, Salisbury, UK under the number 88042101. The mAb has been successfully used to localise the angiogenic blood

vessels of tumours without crossreactivity to mature vascular endothelium, illustrating the potential of FN isoforms for vascular targeting using antibodies.

However, there remain certain caveats to the specificity of the BC-1 mAb. The fact that BC-1 does not directly recognize the B+ isoform has raised the question of whether in some tissues, the epitope recognized by BC-1 could be unmasked without the presence of ED-B and therefore lead indirectly to unwanted crossreactivity of BC-1. Furthermore, BC-1 is strictly specific for the human B+ isoform, meaning that studies in animals on the biodistribution and tumour localisation of BC-1 are not possible. Although polyclonal antibodies to recombinant fusion proteins containing the B+ isoform have been produced (Peters et al, 1995, *Cell Adhes. Commun* 3:67-89), they are only reactive with FN which has been treated with N-glycanase to unmask the epitope.

A further general problem with the use of mouse monoclonal antibodies is the human anti-mouse antibody (HAMA) response (Schroff et al (1985) *Cancer Res* 45: 879-885; DeJager et al (1988) *Proc. Am. Assoc. Cancer Res.* 29:377). HAMA responses have a range of effects, from neutralisation of the administered antibody leading to a reduced therapeutic dose, through to allergic responses, serum sickness and renal impairment.

Although polyclonal antisera reactive with recombinant ED-B have been identified (see above), the isolation of mAbs with the same specificity as BC-1 following immunisation of mice has generally proved difficult because human and mouse ED-B proteins show virtually 100% sequence homology. The human protein may therefore look like a self-antigen to the mouse which then does not mount an immune response to it. In fact, in over ten years of intensive research in this field, only a single mAb has been identified with indirect reactivity to the B+ FN isoform (BC-1), with none recognising ED-B directly. It is almost certainly significant that the specificity of BC-1 is for a cryptic epitope exposed as a consequence of ED-B, rather than for part of ED-B itself, which is likely to be absent from mouse FN and therefore not seen as "self" by the immune system of the mouse.

Realisation of the present invention has been achieved using an alternative strategy where prior immunisation with fibronectin or ED-B is not required: antibodies with specificity for the ED-B isoform have been obtained as single chain Fvs (scFvs) from libraries of human antibody variable regions displayed on the surface of filamentous bacteriophage (Nissim et al., 1994; see also WO92/01047, WO92/20791, WO93/06213, WO93/11236, WO93/19172).

We have found using the human antibody phage library that specific scFvs can be isolated both by direct selection on recombinant FN-fragments containing the ED-B domain and on recombinant ED-B itself when these antigens are coated onto a solid surface ("panning"). These same sources of antigen have also been successfully used to produce "second generation" scFvs with improved properties relative to the parent clones in a process of "affinity maturation". We have found that the isolated scFvs react strongly and specifically with the B+isoform of human FN without prior treatment with N-glycanase.

In anti-tumour applications the human antibody antigen binding domains provided by the present invention have the advantage of not being subject to the HAMA response. Furthermore, as exemplified herein, they are useful in immunohistochemical analysis of tumour tissue, both *in vitro* and *in vivo*. These and other uses are discussed further herein and are apparent to the person of ordinary skill in the art.

TERMINOLOGY

Specific binding member

This describes a member of a pair of molecules which have binding specificity for one another. The

members of a specific binding pair may be naturally derived or wholly or partially synthetically produced. One member of the pair of molecules has an area on its surface, or a cavity, which specifically binds to and is therefore complementary to a particular spatial and polar organisation of the other member of the pair of molecules. Thus the members of the pair have the property of binding specifically to each other. Examples of types of specific binding pairs are antigen-antibody, biotin-avidin, hormone-hormone receptor, receptor-ligand, enzyme-substrate. This application is concerned with antigen-antibody type reactions.

Antibody

This describes an immunoglobulin whether natural or partly or wholly synthetically produced. The term also covers any polypeptide or protein having a binding domain which is, or is homologous to, an antibody binding domain. These can be derived from natural sources, or they may be partly or wholly synthetically produced. Examples of antibodies are the immunoglobulin isotypes and their isotypic subclasses; fragments which comprise an antigen binding domain such as Fab, scFv, Fv, dAb, Fd; and diabodies.

It is possible to take monoclonal and other

antibodies and use techniques of recombinant DNA technology to produce other antibodies or chimeric molecules which retain the specificity of the original antibody. Such techniques may involve introducing DNA encoding the immunoglobulin variable region, or the complementarity determining regions (CDRs), of an antibody to the constant regions, or constant regions plus framework regions, of a different immunoglobulin. See, for instance, EP-A-184187, GB 2188638A or EP-A-239400. A hybridoma or other cell producing an antibody may be subject to genetic mutation or other changes, which may or may not alter the binding specificity of antibodies produced.

As antibodies can be modified in a number of ways, the term "antibody" should be construed as covering any specific binding member or substance having a binding domain with the required specificity. Thus, this term covers antibody fragments, derivatives, functional equivalents and homologues of antibodies, including any polypeptide comprising an immunoglobulin binding domain, whether natural or wholly or partially synthetic. Chimeric molecules comprising an immunoglobulin binding domain, or equivalent, fused to another polypeptide are therefore included. Cloning and expression of chimeric antibodies are described in EP-A-0120694 and EP-A-0125023.

It has been shown that fragments of a whole antibody can perform the function of binding antigens. Examples of binding fragments are (i) the Fab fragment consisting of VL, VH, CL and CH1 domains; (ii) the Fd fragment consisting of the VH and CH1 domains; (iii) the Fv fragment consisting of the VL and VH domains of a single antibody; (iv) the dAb fragment (Ward, E.S. et al., Nature 341, 544-546 (1989)) which consists of a VH domain; (v) isolated CDR regions; (vi) F(ab')2 fragments, a bivalent fragment comprising two linked Fab fragments (vii) single chain Fv molecules (scFv), wherein a VH domain and a VL domain are linked by a peptide linker which allows the two domains to associate to form an antigen binding site (Bird et al, Science, 242, 423-426, 1988; Huston et al, PNAS USA, 85, 5879-5883, 1988); (viii) bispecific single chain Fv dimers (PCT/US92/09965) and (ix) "diabodies", multivalent or multispecific fragments constructed by gene fusion (WO94/13804; P. Holliger et al Proc. Natl. Acad. Sci. USA 90 6444-6448, 1993).

Diabodies are multimers of polypeptides, each polypeptide comprising a first domain comprising a binding region of an immunoglobulin light chain and a second domain comprising a binding region of an immunoglobulin heavy chain, the two domains being linked (e.g. by a peptide linker) but unable to associate with

each other to form an antigen binding site: antigen binding sites are formed by the association of the first domain of one polypeptide within the multimer with the second domain of another polypeptide within the multimer (WO94/13804).

Where bispecific antibodies are to be used, these may be conventional bispecific antibodies, which can be manufactured in a variety of ways (Holliger, P. and Winter G. Current Opinion Biotechnol. 4, 446-449 (1993)), eg prepared chemically or from hybrid hybridomas, or may be any of the bispecific antibody fragments mentioned above. It may be preferable to use scFv dimers or diabodies rather than whole antibodies. Diabodies and scFv can be constructed without an Fc region, using only variable domains, potentially reducing the effects of anti-idiotypic reaction. Other forms of bispecific antibodies include the single chain "Janusins" described in Traunecker et al, Embo Journal, 10, 3655-3659, (1991).

Bispecific diabodies, as opposed to bispecific whole antibodies, may also be particularly useful because they can be readily constructed and expressed in *E.coli*. Diabodies (and many other polypeptides such as antibody fragments) of appropriate binding specificities can be readily selected using phage display (WO94/13804) from libraries. If one arm of the diabody is to be kept

constant, for instance, with a specificity directed against antigen X, then a library can be made where the other arm is varied and an antibody of appropriate specificity selected.

Antigen binding domain

This describes the part of an antibody which comprises the area which specifically binds to and is complementary to part or all of an antigen. Where an antigen is large, an antibody may only bind to a particular part of the antigen, which part is termed an epitope. An antibody binding domain may be provided by one or more antibody variable domains. Preferably, an antigen binding domain comprises an antibody light chain variable region (VL) and an antibody heavy chain variable region (VH).

Specific

This refers to the situation in which one member of a specific binding pair will not show any significant binding to molecules other than its specific binding partner. The term is also applicable where eg an antigen binding domain is specific for a particular epitope which is carried by a number of antigens, in which case the specific binding member carrying the antigen binding domain will be able to bind to the

various antigens carrying the epitope.

Functionally equivalent variant form

This refers to a molecule (the variant) which although having structural differences to another molecule (the parent) retains some significant homology and also at least some of the biological function of the parent molecule, e.g. the ability to bind a particular antigen or epitope. Variants may be in the form of fragments, derivatives or mutants. A variant, derivative or mutant may be obtained by modification of the parent molecule by the addition, deletion, substitution or insertion of one or more amino acids, or by the linkage of another molecule. These changes may be made at the nucleotide or protein level. For example, the encoded polypeptide may be a Fab fragment which is then linked to an Fc tail from another source. Alternatively, a marker such as an enzyme, flourescein, etc, may be linked.

The present invention generally provides a specific binding member which comprises a human antibody antigen binding domain which is specific for the ED-B oncofoetal domain of fibronectin (FN). Specific binding members according to the invention bind the ED-B domain directly. In one embodiment, a specific binding member binds, after treatment of the FN with the protease

thermolysin, to a, any or all FN containing ED-B. In a further embodiment a specific binding member binds to a, any or all FN containing type III homology repeats which include the ED-B domain. Known FNs are identified in two papers by Carnemolla et al., 1989; 1992). Reference to "all FNs containing ED-B" may be taken as reference to all FNs identified in those papers as containing ED-B.

The specific binding member preferably binds human ED-B, and preferably B+FN of at least one other species, such as mouse, rat and/or chicken. Preferably, the specific binding pair member is able to bind both human fibronectin ED-B and a non-human fibronectin ED-B, such as that of a mouse, allowing for testing and analysis of the sbp member in an animal model.

Specific binding pair members according to the present invention bind fibronectin ED-B without competing with the publicly available deposited antibody BC-1 discussed elsewhere herein. BC-1 is strictly specific for human B+ isoform. Specific binding pair members according to the present invention do not bind the same epitope as BC-1.

Binding of a specific binding member according to the present invention to B+FN may be inhibited by the ED-B domain.

In an aspect of the present invention the binding

domain has, when measured as a purified monomer, a dissociation constant (Kd) of 6×10^{-8} M or less for ED-B FN.

In an aspect of the present invention the binding domain is reactive with, i.e. able to bind, fibronectin ED-B without prior treatment of the fibronectin ED-B with N-glycanase.

Specific binding pair members according to the present invention may be provided as isolates or in purified form, that is to say in a preparation or formulation free of other specific binding pair members, e.g. antibodies or antibody fragments, or free of other specific binding pair members able to bind fibronectin ED-B. They may be "monoclonal" in the sense of being from a single clone, rather than being restricted to antibodies obtained using traditional hybridoma technology. As discussed, specific binding pair members according to the present invention may be obtained using bacteriophage display technology and/or expression in recombinant, e.g. bacterial, host cells. There is no prior disclosure of a monoclonal specific binding pair member which directly binds fibronectin ED-B.

The specific binding member may comprise a polypeptide sequence in the form of an antibody fragment such as single chain Fv (scFv). Other types of antibody fragments may also be utilised such as Fab, Fab',

F(ab')2, Fabc, Facb or a diabody (G. Winter and C. Milstein Nature 349, 293-299,1991; WO94/13804). The specific binding member may be in the form of a whole antibody. The whole antibody may be in any of the forms of the antibody isotypes eg IgG, IgA, IgD, IgE and IgM and any of the forms of the isotype subclasses eg IgG1 or IgG4.

The specific binding member may also be in the form of an engineered antibody e.g. a bispecific antibody molecule (or a fragment such as F(ab')2) which has one antigen binding arm (i.e. specific domain) against fibronectin ED-B as disclosed and another arm against a different specificity, or a bivalent or multivalent molecule.

In addition to antibody sequences, the specific binding member may comprise other amino acids, e.g. forming a peptide or polypeptide, or to impart to the molecule another functional characteristic in addition to ability to bind antigen. For example, the specific binding member may comprise a label, an enzyme or a fragment thereof and so on.

The binding domain may comprise part or all of a VH domain encoded by a germ line segment or a re-arranged gene segment. The binding domain may comprise part or all of a VL kappa domain or a VL lambda domain.

The binding domain may comprise a VH1, VH3 or VH4

germ-line gene sequence, or a re-arranged form thereof.

A specific binding member according to the present invention may comprise a heavy chain variable region ("VH" domain) derived from human germline DP47, the sequence of which is shown in Figure 1(a), residues 1 to 98. The 'DP' nomenclature is described in Tomlinson I.M. et al, (1992) J. Mol. Biol. 227: 776-798. The amino acid sequence of the CDR3 may be Ser Leu Pro Lys. The amino acid sequence of the CDR3 may be Gly Val Gly Ala Phe Arg Pro Tyr Arg Lys His Glu. Thus, the binding domain of a specific binding member according to the present invention may include a VH domain that comprises the amino acid sequences shown in Figure 1(a) for CGS1 and CGS2.

The binding domain may comprise a light chain variable region ("VL" domain) derived from human germline DPL16, the sequence of which is shown in Figure 1(b) as codons 1-90.

The VL domain may comprise a CDR3 sequence Asn Ser Ser Pro Val Val Leu Asn Gly Val Val. The VL domain may comprise a CDR 3 sequence Asn Ser Ser Pro Phe Glu His Asn Leu Val Val.

Specific binding members of the invention may comprise functionally equivalent variants of the sequences shown in Figure 1, e.g. one or more amino acids has been inserted, deleted, substituted or added,

provided a property as set out herein is retained. For instance, the CDR3 sequence may be altered, or one or more changes may be made to the framework regions, or the framework may be replaced with another framework region or a modified form, provided the specific binding member binds ED-B.

One or more CDR's from a VL or VH domain of an antigen binding domain of an antibody disclosed herein may be used in so-called "CDR-grafting" in which one or more CDR sequences of a first antibody is placed within a framework of sequences not of that antibody, e.g. of another antibody, as disclosed in EP-B-0239400. CDR sequences for CGS1 and CGS2 are shown in Figure 1(a) and 1(b).

A specific binding member according to the invention may be one which competes with an antibody or scFv described herein for binding to fibronectin ED-B. Competition between binding members may be assayed easily *in vitro*, for example by tagging a specific reporter molecule to one binding member which can be detected in the presence of other untagged binding member(s), to enable identification of specific binding members which bind the same epitope or an overlapping epitope.

A specific binding member according to the present invention may be used in a method comprising causing or

allowing binding of the specific binding member to its epitope. Binding may follow administration of the specific binding member to a mammal, e.g. human or rodent such as mouse.

The present invention provides the use of a specific binding member as above to use as a diagnostic reagent for tumours. Animal model experimental evidence described below shows that binding members according to the present invention are useful in *in vivo* tumour localisation.

Preferred specific binding members according to the present invention include those which bind to human tumours, e.g. in a cryostat section, which show an invasive and angiogenic phenotype and those which bind to embryonic tissues, e.g. in a cryostat section. Binding may be demonstrated by immunocytochemical staining.

In a preferred embodiment, the specific binding member does not bind, or does not bind significantly, tenascin, an extracellular matrix protein.

In another preferred embodiment, the specific binding member does not bind, or does not bind significantly, normal human skin, e.g. in a cryostat section and/or as demonstrated using immunocytochemical staining.

Further embodiments of specific binding members

according to the present invention do not bind, or do not bind significantly, to one or more normal tissues (e.g. in cryostat section and/or as demonstrated using immunocytochemical staining) selected from liver, spleen, kidney, stomach, small intestine, large intestine, ovary, uterus, bladder, pancreas, suprarenal glands, skeletal muscle, heart, lung, thyroid and brain.

A specific binding member for ED-B may be used as an *in vivo* targeting agent which may be used to specifically demonstrate the presence and location of tumours expressing or associated with fibronectin ED-B. It may be used as an imaging agent. The present invention provides a method of determining the presence of a cell or tumour expressing or associated with fibronectin ED-B expression, the method comprising contacting cells with a specific binding member as provided and determining the binding of the specific binding member to the cells. The method may be performed *in vivo*, or *in vitro* on a test sample of cells removed from the body.

The reactivities of antibodies on a cell sample may be determined by any appropriate means. Tagging with individual reporter molecules is one possibility. The reporter molecules may directly or indirectly generate detectable, and preferably measurable, signals. The linkage of reporter molecules may be directly or

indirectly, covalently, eg via a peptide bond or non-covalently. Linkage via a peptide bond may be as a result of recombinant expression of a gene fusion encoding antibody and reporter molecule.

One favoured mode is by covalent linkage of each antibody with an individual fluorochrome, phosphor or laser dye with spectrally isolated absorption or emission characteristics. Suitable fluorochromes include fluorescein, rhodamine, phycoerythrin and Texas Red. Suitable chromogenic dyes include diaminobenzidine.

Other reporters include macromolecular colloidal particles or particulate material such as latex beads that are coloured, magnetic or paramagnetic, and biologically or chemically active agents that can directly or indirectly cause detectable signals to be visually observed, electronically detected or otherwise recorded. These molecules may be enzymes which catalyse reactions that develop or change colours or cause changes in electrical properties, for example. They may be molecularly excitable, such that electronic transitions between energy states result in characteristic spectral absorptions or emissions. They may include chemical entities used in conjunction with biosensors. Biotin/avidin or biotin/streptavidin and alkaline phosphatase detection systems may be employed.

The mode of determining binding is not a feature of the present invention and those skilled in the art are able to choose a suitable mode according to their preference and general knowledge.

The signals generated by individual antibody-reporter conjugates may be used to derive quantifiable absolute or relative data of the relevant antibody binding in cell samples (normal and test). In addition, a general nuclear stain such as propidium iodide may be used to enumerate the total cell population in a sample, allowing the provision of quantitative ratios of individual cell populations relative to the total cells. When a radionucleotide such as ^{125}I, ^{111}In or ^{99m}Tc is attached to an antibody, if that antibody localises preferentially in tumour rather than normal tissues, the presence of radiolabel in tumour tissue can be detected and quantitated using a gamma camera. The quality of the tumour image obtained is directly correlated to the signal:noise ratio.

The antibodies may be utilised as diagnostic agents to trace newly vascularised tumours, and may also be employed, e.g. in modified form, to deliver cytotoxic agents or to trigger coagulation within new blood vessels, thus starving the developing tumour of oxygen and nutrients and constituting an indirect form of tumour therapy.

The present invention also provides for the use of a specific binding member as above to use as a therapeutic reagent, for example when coupled, bound or engineered as a fusion protein to possess an effector function. A specific binding member according to the present invention may be used to target a toxin, radioactivity, T-cells, killer cells or other molecules to a tumour expressing or associated with the antigen of interest.

Accordingly, further aspects of the invention provide methods of treatment comprising administration of a specific binding member as provided, pharmaceutical compositions comprising such a specific binding member, and use of such a specific binding member in the manufacture of a medicament for administration, for example in a method of making a medicament or pharmaceutical composition comprising formulating the specific binding member with a pharmaceutically acceptable excipient.

In accordance with the present invention, compositions provided may be administered to individuals. Administration is preferably in a "therapeutically effective amount", this being sufficient to show benefit to a patient. Such benefit may be at least amelioration of at least one symptom. The actual amount administered, and rate and time-course

of administration, will depend on the nature and severity of what is being treated. Prescription of treatment, eg decisions on dosage etc, is within the responsibility of general practioners and other medical doctors. Appropriate doses of antibody are well known in the art; see Ledermann J.A. et al. (1991) Int J. Cancer 47: 659-664; Bagshawe K.D. et al. (1991) Antibody, Immunoconjugates and Radiopharmaceuticals 4: 915-922.

A composition may be administered alone or in combination with other treatments, either simultaneously or sequentially dependent upon the condition to be treated.

Pharmaceutical compositions according to the present invention, and for use in accordance with the present invention, may comprise, in addition to active ingredient, a pharmaceutically acceptable excipient, carrier, buffer, stabiliser or other materials well known to those skilled in the art. Such materials should be non-toxic and should not interfere with the efficacy of the active ingredient. The precise nature of the carrier or other material will depend on the route of administration, which may be oral, or by injection, e.g. intravenous.

Pharmaceutical compositions for oral administration may be in tablet, capsule, powder or liquid form. A

tablet may comprise a solid carrier such as gelatin or an adjuvant. Liquid pharmaceutical compositions generally comprise a liquid carrier such as water, petroleum, animal or vegetable oils, mineral oil or synthetic oil. Physiological saline solution, dextrose or other saccharide solution or glycols such as ethylene glycol, propylene glycol or polyethylene glycol may be included.

For intravenous, injection, or injection at the site of affliction, the active ingredient will be in the form of a parenterally acceptable aqueous solution which is pyrogen-free and has suitable pH, isotonicity and stability. Those of relevant skill in the art are well able to prepare suitable solutions using, for example, isotonic vehicles such as Sodium Chloride Injection, Ringer's Injection, Lactated Ringer's Injection. Preservatives, stabilisers, buffers, antioxidants and/or other additives may be included, as required.

A specific binding member according to the present invention may be made by expression from encoding nucleic acid. Nucleic acid encoding any specific binding member as provided itself forms an aspect of the present invention, as does a method of production of the specific binding member which method comprises expression from encoding nucleic acid therefor. Expression may conveniently be achieved by culturing

under appropriate conditions recombinant host cells containing the nucleic acid.

The nucleic acid may encode any of the amino acid sequences of the antibody antigen binding domains described herein or any functionally equivalent form. Changes may be made at the nucleotide level by addition, substitution, deletion or insertion of one or more nucleotides, which changes may or may not be reflected at the amino acid level, dependent on the degeneracy of the genetic code.

Systems for cloning and expression of a polypeptide in a variety of different host cells are well known. Suitable host cells include bacteria, mammalian cells, yeast and baculovirus systems. Mammalian cell lines available in the art for expression of a heterologous polypeptide include Chinese hamster ovary cells, HeLa cells, baby hamster kidney cells and many others. A common, preferred bacterial host is *E. coli*.

The expression of antibodies and antibody fragments in prokaryotic cells such as *E. coli* is well established in the art. For a review, see for example Plückthun, A. Bio/Technology 9: 545-551 (1991). Expression in eukaryotic cells in culture is also available to those skilled in the art as an option for production of a specific binding member, see for recent reviews, for example Reff, M.E. (1993) Curr. Opinion Biotech. 4: 573-

576; Trill J.J. et al. (1995) Curr. Opinion Biotech 6: 553-560.

Suitable vectors can be chosen or constructed, containing appropriate regulatory sequences, including promoter sequences, terminator sequences, polyadenylation sequences, enhancer sequences, marker genes and other sequences as appropriate. Vectors may be plasmids, viral e.g. 'phage, or phagemid, as appropriate. For further details see, for example, *Molecular Cloning: a Laboratory Manual*: 2nd edition, Sambrook et al., 1989, Cold Spring Harbor Laboratory Press. Many known techniques and protocols for manipulation of nucleic acid, for example in preparation of nucleic acid constructs, mutagenesis, sequencing, introduction of DNA into cells and gene expression, and analysis of proteins, are described in detail in *Short Protocols in Molecular Biology*, Second Edition, Ausubel et al. eds., John Wiley & Sons, 1992. The disclosures of Sambrook et al. and Ausubel et al. are incorporated herein by reference.

Thus, a further aspect of the present invention provides a host cell containing nucleic acid as disclosed herein. A still further aspect provides a method comprising introducing such nucleic acid into a host cell. The introduction may employ any available technique. For eukaryotic cells, suitable techniques

may include calcium phosphate transfection, DEAE-Dextran, electroporation, liposome-mediated transfection and transduction using retrovirus or other virus, e.g. vaccinia or, for insect cells, baculovirus. For bacterial cells, suitable techniques may include calcium chloride transformation, electroporation and transfection using bacteriophage.

The introduction may be followed by causing or allowing expression from the nucleic acid, e.g. by culturing host cells under conditions for expression of the gene.

In one embodiment, the nucleic acid of the invention is integrated into the genome (e.g. chromosome) of the host cell. Integration may be promoted by inclusion of sequences which promote recombination with the genome, in accordance with standard techniques.

Following production of a specific binding member it may be used for example in any of the manners disclosed herein, such as in the formulation of a pharmaceutical or a diagnostic product, such as a kit comprising in addition to the specific binding member one or more reagents for determining binding of the member to cells, as discussed.

Further aspects of the invention and embodiments

will be apparent to those skilled in the art. In order that the present invention is fully understood, the following examples are provided by way of exemplification only and not by way of limitation.

Reference is made to the following figures:

Figure 1 shows aligned amino acid sequences of the VH and VL of scFvs CGS-1 and CGS-2. Figure 1(a) shows VH sequences; Figure 1(b) shows VL sequences. CDRs (1, 2 and 3) are indicated. The most homologous human germline VH to both scFvs is the DP47 segment of the VH3 family; the VL segment of both clones is DPL16, the light chain used to build the original scFv library (Nissim et al, 1994). Residues that distinguish the two clones from each other are underlined.

Figure 2: Figure 2A shows a model of the domain structure of a human FN subunit. The IIICS, ED-A and ED-B regions of variability, due to alternative splicing of the FN pre-mRNA, are indicated. The figure also indicates the internal homologies as well as the main thermolysin digestion products containing ED-B (Zardi et al, 1987). Figure 2B shows 4-18% SDS-PAGE of plasma and WI38VA FN and their thermolysin digests stained with Coomassie Blue and immunoblots probed with BC-1, IST-6, CGS-1 and CGS-2. Undigested (lane 1) and digested plasma FN using thermolysin at 1 μg/mg of FN (lane 3) and 10 μg/mg of FN (lane 4). Undigested (lane 2) and

digested WI38VA FN using thermolysin at 1μg/mg (lane 5), 5μg/mg (lane 6) and 10μg/mg (lane 7) of FN. The numbers on the right hand side indicate the main thermolysin digestion products shown in Figure 2A. The values on the left indicate the molecular weight standards in kiloDalton (kD).

Figure 3: Figure 3A shows the FN type III repeat sequences contained in the fusion and recombinant proteins expressed in E. coli and the reactivity of these proteins with CGS-1 and CGS-2 and with the mAbs BC-1 and IST-6. Figure 3B shows a Coomassie Blue stained gel and alongside the immunoblots probed with CGS-1, CGS-2, BC-1, IST-6. The numbering of the lanes corresponds to that of the peptide constructs in the upper part of the figure. The values on the left indicate the molecular weight standards in kD.

All documents mentioned herein are incorporated by reference.

List of Examples

Example 1 - Isolation of human scFvs specific for the ED-B domain of human FN.

Example 2 - Affinity maturation of human scFvs specific for the ED-B domain of human FN.

Example 3 - Specificity of affinity matured scFvs

for ED-B-containing fibronectins.

Example 4 - The use of affinity matured anti-ED-B scFvs in immunocytochemical staining of human and mouse tumour sections.

Example 5 - The use of affinity matured anti-ED-B scFvs in *in vivo* targeting of human tumours.

EXAMPLE 1 - Isolation of human scFvs specific for the ED-B domain of human FN

To prepare the recombinant FN peptides containing the type III repeats 2-11 (B-) and 2-11 (B+), a construct was made using FN cDNA from the clones pFH154 (Kornblihtt et al 1985), IF10 and IF2 (Carnemolla et al, 1989). The cDNA constructs, spanning bases 2229-4787, (Kornblihtt et al, 1985) were inserted into the vector pQE-3/5 using the QIAexpress kit from Qiagen (Chatsworth, CA). The recombinants FN-III 2-11 (B-) and (B+) were purified by immunoaffinity chromatography using the mAb 3E3 (Pierschbacher et al 1981) conjugated to Sepharose 4B (Pharmacia). DNA fragments for the preparation of the recombinant FN fragments containing the type III homology repeats 7B89, 789, ED-B and FN-6 were produced by polymerase chain reaction (PCR) amplification using UltMa DNA polymerase (Perkin Elmer), using cDNA from clones FN 2-11 (B+) and FN 2-11 (B-) as template. Primers were designed to allow cloning of PCR

products into pQE-12 using the QIAexpress kit (Qiagen) and transformation into E.coli. All cDNA clones were sequenced using a Sequenase 2.0 DNA sequencing kit (USB).

Recombinant proteins were purified by Ni-NTA chromatography (IMAC), according to the manufacturers' instructions (Qiagen), using the hexahistidine tag at the carboxy terminus of the FN fragments. The ED-B-βGal fusion protein was prepared by cloning ED-B cDNA into the λgt11 bacteriophage vector, to give clone λED-B. Clone λchFN60 (containing part of the ED-B sequence) was derived as a fusion protein from the cloned chicken FN pchFN60 (Norton et al, 1987).

Two different forms of the ED-B isoform were used as a source of antigen for the selection of the human scFv library, and in both cases, the isoform was recombinant human protein. Three rounds of panning were performed after coating each antigen onto immunotubes (Nunc; Maxisorp) overnight at 50 μg/ml in PBS (20mM phosphate buffer, 0.15M NaCl, pH 7.2). The first antigen was the recombinant FN fragment 7B89, in which the ED-B domain is flanked by the adjacent type III FN homology repeats; this was coated at 4°C overnight. The second antigen used was recombinant ED-B (Zardi et al, 1987) with a carboxy terminal hexahistidine tag; this protein does not contain lysine residues, so that the

terminal amino group of the first amino acid is available for site-specific covalent immobilisation of ED-B to reactive ELISA plates (Nunc; Covalink). Coating was carried out overnight at room temperature.

After three rounds of panning, the eluted phage were infected into HB2151 E.coli cells and plated as described (Nissim et al., 1994). After each round of selection, 95 ampicillin-resistant single colonies were screened to identify antigen-specific scFvs by ELISA. Clones which gave the highest ELISA signals on the antigens used for panning were selected for further analysis and for affinity maturation. These clones were also demonstrated to give specific staining of sections of glioblastoma multiforme and breast tumours by immunocytochemical staining, described in more detail in Example 4.

Example 2 - Affinity maturation of human scFvs specific for the ED-B domain of human FN

Clones 35GE (from selection with 7B89) and 28SI (from selection with the ED-B domain alone) were selected as candidate antibodies for affinity maturation. In order to diversify the light chains as a means of improving affinity, six amino acid positions of the VL CDR3 of each antibody were randomised and the resulting libraries from each clone were displayed on

phage.

In brief, a single colony expressing the relevant antibody was PCR amplified with primers LMB3 (5' CAG GAA ACA GCT ATG AC 3') and CDR3-6-VL-FOR (5' CTT GGT CCC TCC GCC GAA TAC CAC MNN MNN MNN MNN MNN MNN AGA GGA GTT ACA GTA ATA GTC AGC CTC 3') (94°C [1'] - 55°C [1'] - 72°C [1'30"], 25 cycles). The resulting product was gel-purified and used as template for a second amplification step with primers LMB3 and Jλ-Not-FOR (5' ATT GCT TTT CCT TTT TGC GGC CGC GCC TAG GAC GGT CAG CTT GGT CCC TCC GCC 3') (94°C [1'] - 55°C [1'] - 72°C [1'30"], 25 cycles). The crude PCR product, which ran as a single band of the correct molecular weight on an agarose gel, was directly purified from the PCR mixture using Spin-Bind (FMC, Rockland, ME, USA), double-digested with the restriction enzymes Nco1/Not1 and ligated into Nco1/Not1-digested pHEN1 (Hoogenboom et al., 1991). Approximately 5 μg of vector and of insert were used in the ligation mix, which was extracted once with phenol, once with phenol/chloroform/isoamyl alcohol (25:24:1), then ethanol-precipitated in the presence of glycogen (Boehringer, Mannheim, Germany) as a carrier and dried in a vacuum desiccator. The pellet was resuspended in 20 μl water and transformed into 100 μl aliquots of electrocompetent TGI *E. coli* cells (Gibson, 1984), yielding a library of > 10^6 individual clones.

The maturation library was then processed as for the Nissim library (Nissim et al., 1994) to produce phage particles, which were used for one round of selection on immunotubes using 7B89 (10 μg/ml) as antigen, followed by a round of kinetic selection (Hawkins et al., 1992). This selection step was performed by incubating biotinylated 7B89 (10 nM) with the phage suspension (approx. 10^{12} t.u.) in 2% milk-PBS (2% MPBS) from the first round of selection for 5 minutes, then adding non-biotinylated 7B89 (1 μM) and letting the competition proceed for 30 minutes. 100 μl of streptavidin-coated dynabeads (Dynal: M480) preblocked in 2% MPBS were then added to the reaction mixture, mixed for 2 minutes and then captured on a magnet and washed 10 times with alternate washes of (PBS + 0.1% Tween-20) and PBS. Phage were eluted from the beads with 0.5 ml 100 mM triethylamine. This solution was then neutralised with 0.25 ml 1M Tris, pH 7.4, and used to infect exponentially growing HB2151 cells (Nissim et al., 1994). 95 ampicillin-resistant single colonies were used to produce scFv-containing supernatants (Nissim et al., 1994) which were screened by ELISA, immunohistochemistry and BIAcore to identify the best binders. They were then subcloned between Sfi1/Not1 sites of the pDN268 expression vector (Neri et al., 1996), which appends a phosphorylatable tag, the

FLAG epitope and a hexahistidine tag at the C-terminal extremity of the scFv.

Single colonies of the relevant antibodies subcloned in pDN268 were grown at 37°C in 2xTY containing 100 mg/l ampicillin and 0.1% glucose. When the cell culture reached OD^{600} = 0.8, IPTG was added to a final concentration of 1 mM and growth continued for 16-20 hrs at 30°C. After centrifugation (GS-3 Sorvall rotor, 7000 rpm, 30 minutes), the supernatant was filtered, concentrated and exchanged into loading buffer (50 mM phosphate, pH 7.4 500 mM NaCl, 20 mM imidazole) using a Minisette (Filtron) tangential flow apparatus. The resulting solution was loaded onto 1 ml Ni-NTA resin (Qiagen), washed with 50 ml loading buffer and eluted with elution buffer (50mM phosphate, pH 7.4, 500mM NaCl, 100 mM imidazole). The purified antibody was analysed by SDS-PAGE (Laemmli, 1970) and dialysed versus PBS at 4°C. Purified scFv preparations were further processed by gel-filtration using an FPLC apparatus equipped with a S-75 column (Pharmacia), since it is known that multivalent scFv fragments may exhibit an artificially good binding on BIAcore (Jonsson et al., 1991) by virtue of avidity effects (Nissim et al., 1994; Crothers and Metzger, 1972). The antibody concentration of FPLC-purified monomeric fractions was determined spectrophotometrically assuming an absorbance at 280 nm

of 1.4 units for a 1 mg/ml scFv solution.

Binding of monovalent scFv at various concentrations in the 0.1 - 1 μM range in PBS was measured on a BIAcore machine (Pharmacia Biosensor), using the following antigens: (i) 1000 Resonance Units (RU) of biotinylated recombinant FN fragment 7B89 immobilised on a streptavidin coated chip, which was bound specifically by 250 RU of scFv; (ii) 200 RU of recombinant ED-B, chemically immobilised at the N-terminal amino group, which was bound specifically by 600 RU of scFv; (iii) 3500 RU of ED-B-rich fibronectin WI38VA (see Example 3), which was bound specifically by 150 RU of scFv. Kinetic analysis of the data was performed according to the manufacturers' instructions.

On the basis of qualitative BIAcore analysis of antibody-containing supernatants, one affinity-matured version of each scFv clone was selected: clone CGS-1 from selection with the 78B9 fragment and CGS-2 from selection with ED-B recombinant FN fragment. The association rate constants (kon) and dissociation rate constants (koff) are shown in Table 1, together with the calculated equilibrium dissociation constants (Kd) of both scFvs. Although both clones have Kds in the nanomolar range, clone CGS-2 showed the best improvement over its parent clone, giving a Kd of ~1-2nM (improved from 100nM) with respect to all three proteins tested on

the sensor chip (Table 1).

The affinity matured clones CGS-1 and CGS-2 were sequenced and aligned to a database of human germline antibody V genes (V-BASE) then translated using MacVector software. The VH gene of both clones was most homologous to human germline DP47 (VH3), and in addition each clone had a different VH CDR3 sequence (Figure 1). The VL gene of both clones was the DPL16 germline used in the construction of the human synthetic scFv repertoire described in Nissim et al, 1994. The VL CDR3 sequences differed from each other at four out of six of the randomised residues (Figure 1b).

Example 3 - Specificity of affinity matured scFvs for ED-B-containing fibronectins

The immunoreactivity of the two affinity matured scFvs, CGS-1 and CGS-2, was assessed initially by ELISA and compared directly to the mAb BC-1 (which recognises the B-FN isoform) and mAb IST-6, which only recognises FN isoforms lacking ED-B (Carnemolla et al., 1989; 1992). The characterisation of these mAbs has been previously reported (Carnemolla et al, 1989; 1992). Fine specificity analysis was thereafter carried out using an extensive panel of FN fragments derived by thermolysin treatment and of recombinant fusion proteins.

The antigens used for ELISA and immunoblotting were prepared as follows. FN was purified from human plasma and from the conditioned medium of the WI38VA13 cell line as previously reported (Zardi et al, 1987). Purified FNs were digested with thermolysin (protease type X; Sigma Chemical Co.) as reported by Carnemolla et al (1989). Native FN 110kD (B-) and native FN 120kD (B+) fragments (see Figure 2) were purified from a FN digest as previously reported (Borsi et al, 1991). The large isoform of tenascin-C was purified as reported by Saginati et al (1992). Recombinant proteins were expressed and purified as described in Example 1. SDS-PAGE and Western blotting were carried out as described by Carnemolla et al (1989).

All antigens used in ELISA were diluted in PBS to between 50-100 μg/ml and coated at 4°C overnight onto Immuno-Plate wells (Nunc, Roskilde, Denmark). Unbound antigen was removed with PBS and plates were then blocked with PBS containing 3% (w/v) bovine serum albumin (BSA) for 2h at 37°C. This was followed by four washes with PBS containing 0.05% Tween 20 (PBST). Antibodies were then allowed to bind at 37°C for 1.5h; scFvs were preincubated with an antiserum directed against the tag sequence: mAb M2 [Kodak, New Haven CT] for the FLAG tag or 9E10 [ATCC, Rockville, MD] for the myc tag. Control antibodies tested were mAbs BC-1 and

IST-6. After four washes with PBST, the plates were incubated for 1h at 37°C with 1:2000 diluted (in PBST+3% BSA) biotinylated goat anti-mouse IgG (Bio-SPA Division, Milan, Italy). The washes were repeated and Streptavidin-biotinylated alkaline phosphatase complex (Bio-SPA Division, Milan, Italy) was added (1:800 diluted in PBST containing 2mM MgCl2) for 1h at 37°C. The reaction was developed using Phosphatase substrate tablets (Sigma) in 10% diethanolamine, pH 9.8 and the optical density was read at 405nm. The results are presented in Table 2.

Both CGS-1 and CGS-2 recognised the recombinant ED-B peptide, as well as all native or recombinant FN fragments containing the ED-B sequence, while they did not bind to any FN fragments lacking ED-B. Furthermore, CGS-1 and CGS-2 did not react with tenascin (which comprises fifteen type III homology repeats: Siri et al, 1991) and plasma FN, which does not contain detectable levels of the ED-B sequence in thermolysin digestion products (Zardi et al, 1987). In contrast, CGS-1 and CGS-2 reacted strongly with FN purified from the SV40-transformed cell line WI38VA. About 70-90% of FN molecules from this cell line contain ED-B, as shown by thermolysin digestion and S1 nuclease experiments using purified FN and total RNA prepared from the cell line (Zardi et al, 1987; Borsi et al, 1992). The specificity

of the scFvs for the ED-B component of FN was demonstrated still further by using soluble recombinant ED-B to inhibit binding of CGS-1 and/or CGS-2 to FN on WI38VA cells (data not shown).

The data confirm that CGS-1 and CGS-2 only react specifically with FN derivatives that contain the ED-B domain. They both show the same reactivity as mAb BC-1, except in the case of recombinant ED-B, which was not recognised by BC-1. The intensity of the ELISA signals obtained relative to the mAb controls reflects the high specificity of the two scFvs for ED-B-containing antigens.

The specificity of CGS-1 and CGS-2 was investigated further on immunoblots using FN from plasma and WI38VA cells, and thermolysin digests thereof. Upon thermolysin digestion, FN from WI38VA cells (the majority of which contains ED-B) generates a 120kD fragment (containing ED-B) and a minor 110kD fragment which lacks ED-B (Figure 2A; Zardi et al, 1987). Further digestion of the 120kD domain generates two fragments: a 85kD fragment which contains almost the entire ED-B sequence at its carboxy terminus, and a 35kD sequence (Figure 2A; Zardi et al, 1987).

On the left hand side of Figure 2B is a Coomassie stained gel of the protein fractions analysed by immunoblotting. Plasma FN (lane 1) and thermolysin

digests of the protein (lane 3, containing the 110kD protein, and lane 4, containing digested 110kD protein) were not recognised by CGS-1 and CGS-2. In contrast, ED-B-rich FN from WI38VA cells, both intact (lane 2) and after increasing thermolysin digestion (lanes 5, 6 and 7) was recognised by both scFv fragments. The smallest FN-derived fragment that could be recognised specifically by CGS-1 was the 120kD protein (spanning type III repeats 2-11 inclusive), while CGS-2 was able to recognise the 85kD fragment spanning repeats 2-7 in addition to the N-terminus of ED-B (Figure 2B; Zardi et al, 1987). These results indicate that the two scFvs are reactive to distinct epitopes within the ED-B sequence. The binding of CGS-2 to the 85kD domain indicates that the epitope for this clone lies in the amino terminus of ED-B. In contrast, the loss of CGS-1 binding when the 120kD domain is digested to 85kD demonstrates that it recognises an epitope located more toward the carboxy terminus of the ED-B molecule.

The fine specificity of CGS-1 and CGS-2 was investigated further by immunoblotting using recombinant FN fragments and fusion proteins with or without the ED-B sequence. The FN fusion proteins were prepared as described by Carnemolla et al (1989). The results of these experiments are shown in Figure 3; for the association of the schematic diagram to the structure of

the domains of human FN, see Carnemolla et al, 1992. The binding profiles obtained essentially confirmed what had previously been found by ELISA and immunoblots on purified FN and proteolytic cleavage products: CGS-1 and CGS-2 were strongly reactive with ED-B-containing FN fragments (lanes 2 and 4) but showed no reactivity to FN sequences lacking ED-B (lanes 1 and 3). CGS-1 did not react with either the human (lane 5) or the chicken (lane 6) ED-B fusion protein, while CGS-2 reacted strongly with both fragments (Figure 3). This result may reflect certain conformational constraints of the epitope in ED-B-containing FN recognised by CGS-1; it is possible, for example, that the epitope is sensitive to denaturation or is not presented correctly when fractinated by SDS-PAGE and transferred to a solid support such as nitrocellulose.

Taken together, these results demonstrate that CGS-1 and CGS-2 bind strongly and specifically to ED-B-containing FNs, at regions distinct from one another and distinct from the ED-B structure which is recognised by the mAb BC-1.

Example 4 - The use of affinity matured anti-ED-B scFvs in immunocytochemical staining of human and mouse tumours

CGS-1 and CGS-2 have both been used to

immunolocalise ED-B containing FN molecules in various normal and neoplastic human tissues. For normal tissue, skin was chosen, since the B-FN isoform is known to be expressed in macrophages and fibroblasts during cutaneous wound healing (Carnemolla et al, 1989; Brown et al, 1993). The two human tumours selected have previously been analysed for the specificity of staining with anti-fibronectin mAbs: glioblastoma multiforme has been studied in detail because endothelial cells in the vessels of this tumour are in a highly proliferative state with increased angiogenetic processes including the expression of B-FN isoforms (Castellani et al, 1994). Furthermore, studies using a diverse panel of normal, hyperplastic and neoplastic human breast tissues have provided further evidence of a correlation between angiogenesis and B-FN expression (Kaczmarek et al, 1994).

For the experiments described here, the immunohistochemical staining of CGS-1 and CGS-2 has been compared to that of mAb BC-1 (which recognises the B-FN isoform) and other mAbs known to react either to all known FN isoform variants (IST-4) or only to FN isoforms lacking ED-B (IST-6). The characterisation of all of these control antibodies has been previously reported (Carnemolla et al, 1989; 1992).

Normal and neoplastic tissues were obtained from

samples taken during surgery. It has already been established that the preparation and fixation of tissues is critical for accurate and sensitive detection of FN-containing molecules (Castellani et al, 1994). For immunohistochemistry, 5μm thick cryostat sections were air dried and fixed in cold acetone for ten minutes. Immunostaining was performed using a streptavidin-biotin alkaline phosphatase complex staining kit (Bio-SPA Division, Milan, Italy) and naphthol-AS-MX-phosphate and Fast Red TR (Sigma). Gill's haematoxylin was used as a counterstain, followed by mounting in glycergel (Dako, Carpenteria, CA) as reported previously by Castellani et al, 1994. In order to analyse specificity further in experiments where positive staining of tissues was obtained, specificity for ED-B was demonstrated by preincubation of antibodies with the recombinant ED-B domain, followed by detection as previously described.

The results of these experiments overall showed that both CGS-1 and CGS-2 reacted with the same histological structures as mAb BC-1. The staining pattern obtained with skin using CGS-1, CGS-2 and BC-1 reflects the absence of ED-B from the FN expressed in the dermis. In the staining of invasive ductal carcinoma sections, CGS-1, CGS-2 and BC-1 showed a restricted distribution of staining, confined to the border between the neoplastic cells and the stroma.

This is consistent with the fact that although total FN is homogeneously distributed throughout the tumour stroma, the expression of B-FN is confined to certain regions, and it is these areas that had previously been successfully localised (in 95% of cases) in invasive ductal carcinoma using mAb BC-1 (Kaczmarek et al, 1994).

Previous findings in the staining of BC-1 of glioblastoma multiforme tumour have been confirmed. Castellani et al (1994) had observed a typical pattern of staining of glomerular-like vascular structures, and in our experiments, CGS-1 and CGS-2 have been shown to give qualitatively identical results.

There is, however, an important difference between CGS-1 and CGS-2 and the mAb BC-1: the two human scFvs have been demonstrated to bind to both chicken and mouse B-FN, whereas BC-1 is strictly human-specific. CGS-2 reacted with chicken embryos (data not shown) and both CGS-1 and CGS-2 reacted with mouse tumours.

CGS-1 staining of vascular structures on sections of the murine F9 teratocarcinoma has also been shown. In contrast, all normal mouse tissues tested (liver, spleen, kidney, stomach, small intestine, large intestine, ovary, uterus, bladder, pancreas, suprarenal glands, skeletal muscle, heart, lung, thyroid and brain) showed a negative staining reaction with CGS-1 and CGS-2 (data not shown). The structures stained in the F9

teratocarcinoma sections were shown to be ED-B specific by using the recombinant ED-B domain to completely inhibit the staining obtained (data not shown).

Example 5 - The use of affinity matured anti-ED-B scFvs in vivo targeting of human tumours

An infrared mouse-imager for immunophotodetection in nude mice (Folli et al., 1994) was built in-house. A 100 W tungsten-halogen lamp equipped with a CY-7 excitation filter (Chroma Technologies, Brattleboro, VT, USA) was used to illuminate a mouse temporarily restrained in a black-painted box. Above the box, an integrating monochrome CCD-camera (Pulnix) with a C-mount lens and a CY-7 emission filter (Chroma Technologies) interfaced with a processing board and computer-controlled by the ImageDok software which enables to specify contrast, gain and integration time (Kinetic Imaging, Liverpool, UK) was used to detect in real-time the antibody-associated CY-7 fluorescence.

Recombinant antibodies were labeled with CY-7-NHS ester (Amersham) according to the manufacturer's instructions. The human melanoma cell-line SKMEL-28 was used to develop xenografted tumours in nude mice, by injecting 1 x 10^7 cells/mouse subcutaneously in one flank. Mice bearing tumours were injected in the tail vein with 100 μl of 1 mg/ml $scFv_1$-$Cy7_1$ solution in PBS,

and imaged at various times with the above described infra-red mouse imager.

These experiments demonstrated that both scFv's localised to the tumour when visualised at a macroscopic level.

Microscopic demonstration of targeting of neovasculature of developing tumours with the two anti-EDB scFvs was detailed as follows.

Nude mice and/or SCID mice bearing either a xenografted SKMEL-28 human melanoma or a mouse F9 teratocarcinoma in one flank, were injected with either unlabeled scFv fragments with the FLAG tag, or biotinylated scFv fragments.

Mice were sacrificed at different time points after injection, tumour and non-tumour sections obtained, which were then stained with conventional immunohistochemistry protocols, using either the anti-FLAG M2 antibody (Kodak, 181) or streptavidin-based detection reagents. Optimal targeting was generally obtained at 12 hours post injection. Both CGS1 and CGS2 wwere demonstrated to bind the neovasculature of both the xenografted human tumour and the murine teratocarcinoma.

Summary

The present invention provides in exemplary

embodiments two scFvs, CGS-1 and CGS-2, which were isolated from the affinity maturation of scFvs isolated from a repertoire of antibody variable regions from unimmunised humans. They have been shown to have dissociation constants for ED-B in the nanomolar range when measured by surface plasmon resonance. As well as binding to different recombinant forms of human ED-B, both scFvs bind to ED-B-containing FN molecules from different species without prior N-glycanase treatment. Furthermore, both CGS-1 and CGS-2 recognised tumour neovasculature, as evinced by immunohistochemistry on tumour cryostat sections.

The antibodies may be utilised in various ways, e.g as diagnostic agents to trace newly vascularised tumours, and may also be modified to deliver cytotoxic agents or to trigger coagulation within new blood vessels, thus starving the developing tumour of oxygen and nutrients and constituting an indirect form of tumour therapy.

TABLE 1

Kinetics and dissociation constants of monomeric scFv fragments CGS-1 and CGS-2 towards ED-B domain-containing proteins.

	rec 7-9 (B+)			rec ED-B			W138VA-FN		
	k_{on}	k_{off}	K_d	k_{on}	k_{off}	K_d	k_{on}	k_{off}	K_d
CGS-1	$1.1x10^5$	$3.9x10^{-3}$	35.5 nM	$1.3x10^5$	$7.0x10^{-3}$	53 nM	$4.1x10^5$	$5.0x10^{-3}$	12.2 nM
CGS-2	$1.1x10^5$	$2.3x10^{-3}$	2.1 nM	$1.3x10^5$	$1.5x10^{-4}$	1.1 nM	$2.9x10^5$	$6.5x10^{-4}$	2.4 nM

TABLE 2

Immunoreactivity of scFv and monoclonal antibodies with fibronectin-derived antigens measured by ELISA. The values represent the OD measured at 405nm after subtraction of background signal. The data are the mean of four experiments showing a maximum 10% standard deviation.

The identity of the different forms of fibronectin used in the experiment is as follows: Plasma FN = human plasma fibronectin; WI38-VA FN = fibronectin from supernatants of SV40-transformed fibroblasts (Zardi et

al, 1987); n110kD = thermolysin treated FN domain 4, without ED-B; n120kD = thermolysin treated FN domain 4, containing ED-B; rec FN7B89 = ED-B domain flanked by adjacent type III FN homology repeats; rec FN789 = type III FN homology repeats with an ED-B domain; rec ED-B = recombinant ED-B alone; rec FN6 = recombinant FN domain 6.

	CGS-1	CGS-2	BC-1	IST-6
Plasma FN	0.07	0.04	0.09	1.73
WI38VA FN	1.16	0.72	1.20	1.12
n110 kD (B-)	0.03	0.01	0.05	1.20
n120 kD (B+)	0.82	0.81	1.20	0.02
rec FN7B89	1.11	1.02	1.02	0.01
rec FN789	0.01	0.01	0.05	1.25
rec ED-B	1.21	1.32	0.15	0.04
rec FN-6	0.01	0.01	0.08	0.03
Tenascin	0.01	0.02	0.06	0.02

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Figure 1:

(a)

	FR1	CDR1	FR2	CDR2
	10 20 30		40	50 60
CGS1	QVQLVESGGGLVQPGGSLRLSCAVSGFTFS	SYAMS	WVRQAPGKGLEWVS	AISGSGGSTYYADSVKG
CGS2	EVQLVESGGGLVQPGGSLRLSCAASGFTFS	SYAMS	WVRQAPGKGLEWVS	AISGSGGSTYYADSVKG

		CDR3	
	70 80 90 98		
CGS1	RFTISRDNSKNTLYLQMNSLRAEDTAVYYCAR	SLPK	WGQGTLVTVSR
CGS2	RFTISRDNSKNTLYLQMNSLRAEDTAVYYCAR	GVGAFRPYRKHE	WGQGTLVTVSR

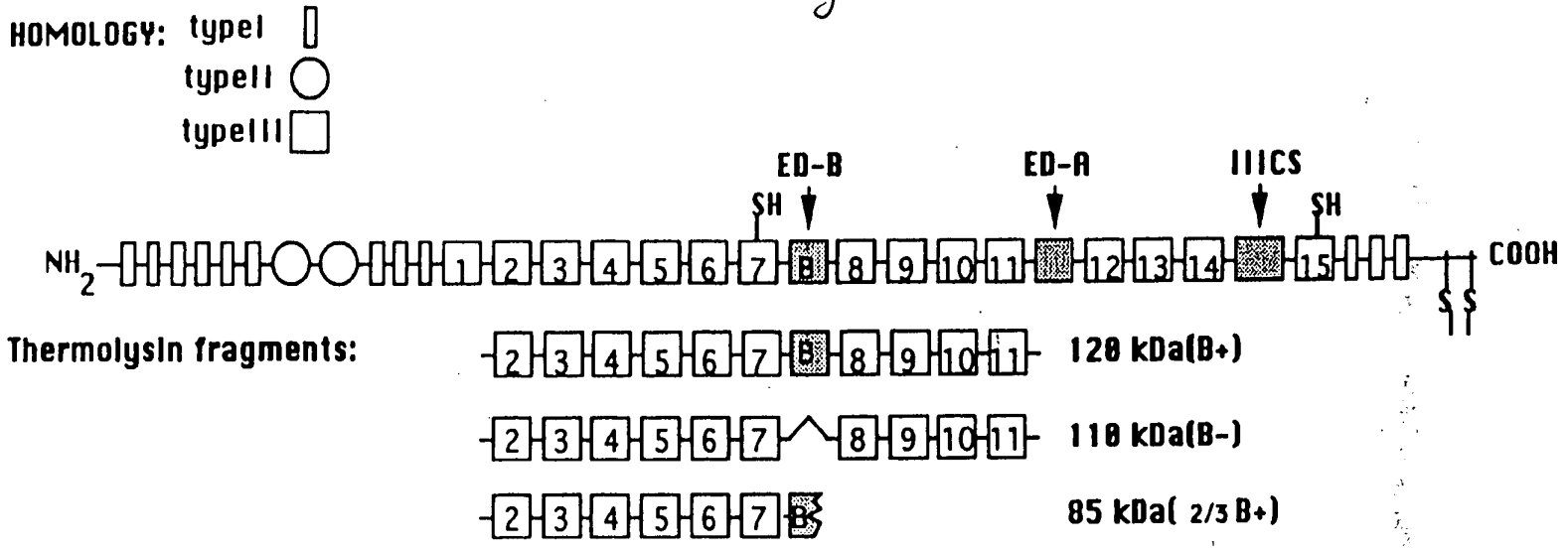
(b)

		CDR1		CDR2
	10 20	30	40	50
CGS1	SSELTQDPAVSVALGQTVRITC	QGDSLRSYYAS	WYQQKPGQAPVLVIY	GKNNRPS
CGS2	SSELTQDPAVSVALGQTVRITC	QGDSLRSYYAS	WYQQKPGQAPVLVIY	GKNNRPS

		CDR3	
	60 70 80	90	100
CGS1	GIPDRFSGSSSGNTASLTITGAQAEDEADYYC	NSSPVVLNGVV	FGGGTKLTVLG
CGS2	GIPDRFSGSSSGNTASLTITGAQAEDEADYYC	NSSPFEHNLVV	FGGGTKLTVLG

A

Figure 2A



B

Figure 2B

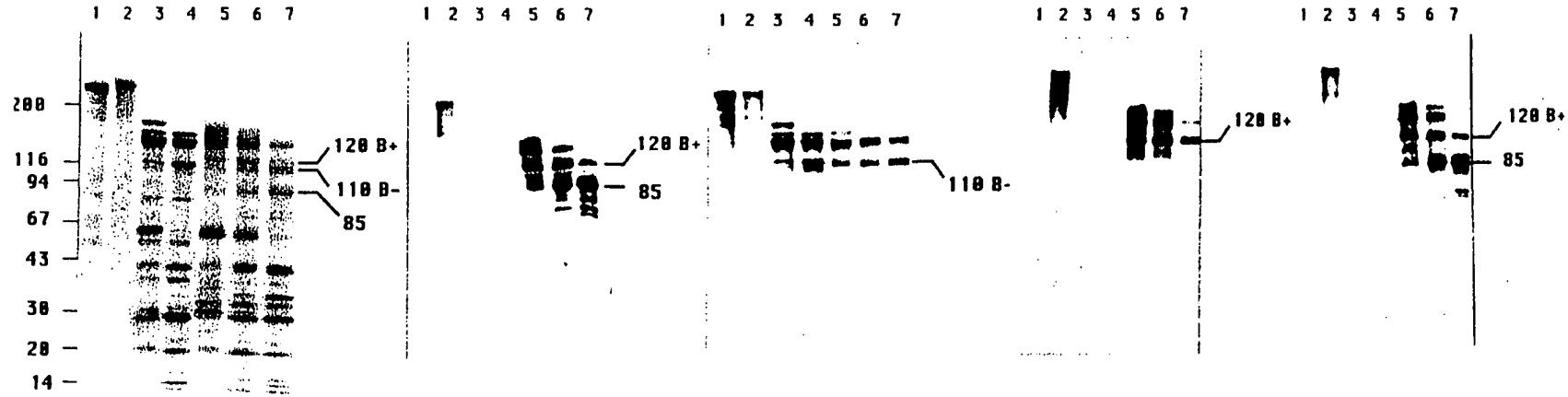
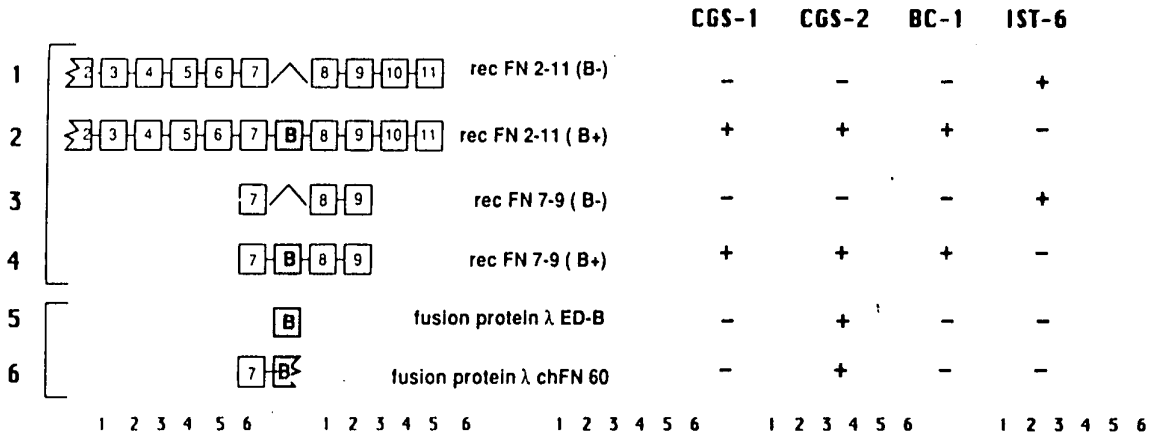


Figure 3A



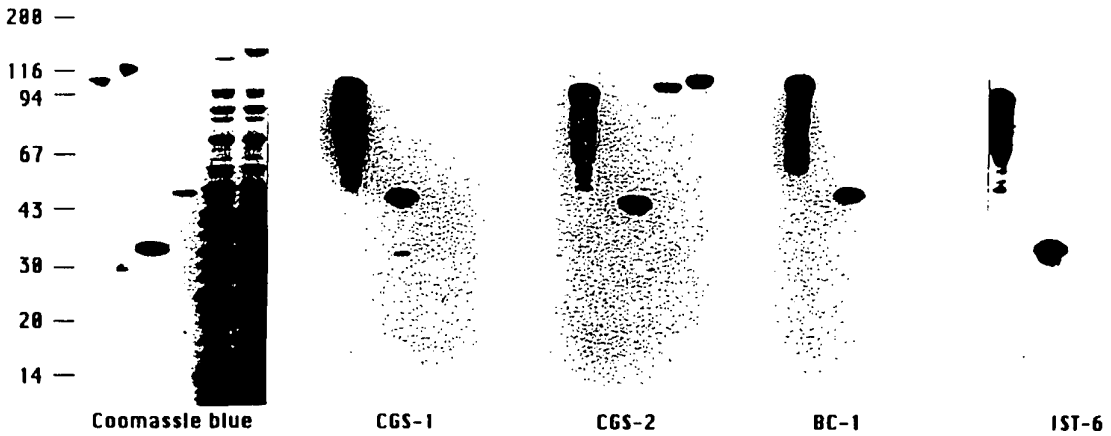


Figure 3B

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